

Title                      Enzyme-aided alkaline extraction of  
                                 oligosaccharides and polymeric xylan from  
                                 hardwood kraft pulp

Author(s)                Hakala, Terhi; Liitiä, Tiina; Suurnäkki, Anna

Citation                   Carbohydrate Polymers. Elsevier.  
                                 Vol. 93 (2013) No: 1, Pages 102 - 108

Date                        2013

URL                        <http://dx.doi.org/10.1016/j.carbpol.2012.05.013>

Rights                     This article may be downloaded for personal  
                                 use only.

**VTT**  
<http://www.vtt.fi>  
P.O. box 1000  
FI-02044 VTT  
Finland

By using VTT Digital Open Access Repository you are bound by the following Terms & Conditions.

I have read and I understand the following statement:

This document is protected by copyright and other intellectual property rights, and duplication or sale of all or part of any of this document is not permitted, except duplication for research use or educational purposes in electronic or print form. You must obtain permission for any other use. Electronic or print copies may not be offered for sale.

1 **Enzyme-aided alkaline extraction of oligosaccharides and polymeric**  
2 **xylan from hardwood kraft pulp**

3 Terhi K. Hakala\*, Tiina Liitiä, and Anna Suurnäkki

4

5 *VTT Technical research centre of Finland, P. O. Box 1000, VTT, Finland*

6 \*Corresponding author: Tel.: +358 20 722 7417, fax: +358 20 722 7071, E-mail: [Terhi.Hakala@VTT.fi](mailto:Terhi.Hakala@VTT.fi)

7

8 **Abstract**

9

10 In this paper we describe the effect of enzyme treatments on the production of  
11 polymeric xylan, oligosaccharides and hemicellulose lean pulp by alkaline extraction of  
12 bleached hardwood kraft pulp. Enzyme treatments were carried out before one or in  
13 between two subsequent alkaline extractions by purified *Trichoderma reesei* xylanase  
14 and endoglucanase II (Cel 5a) as well as by a commercial monocomponent  
15 endoglucanase (FibreCareR). Without enzyme pre-treatment 61% and 7% of the pulp  
16 xylan was extracted in high purity in the first and second alkaline stage, respectively.  
17 Higher molecular mass xylan was obtained in the second than in the first alkaline  
18 extraction. Xylanase treatment before alkaline extraction hydrolysed up to 12% of xylan  
19 to xylooligosaccharides. According to our results, preparation of polymeric xylan,  
20 and/or oligosaccharides as well as hemicellulose lean pulp with cellulose content of 93-  
21 94%, is possible by enzyme-aided alkaline extraction process.

22

23 *Keywords: Enzyme treatment, xylan, xylanase, endoglucanase, oligosaccharide,*  
24 *dissolving pulp*

1

## 2 **1. Introduction**

3

4 Currently the utilization of lignocellulose-based raw materials for novel end-uses is  
5 under vigorous investigation. The lignocellulose biorefinery research is greatly focusing  
6 on the production of biofuels and chemicals derived from abundant biomass resources.  
7 The production of several high value products such as polymeric hemicelluloses and  
8 oligosaccharides in addition to the main product could have a major impact on the  
9 economy of the biorefineries. The polymeric wood hemicelluloses, e.g. xylan and  
10 glucomannan, are interesting starting components for material applications, chemicals  
11 and liquid fuels. For example xylan as such or after modification has end-use applications  
12 in pulp and paper making, food and pharmaceutical industries (Ebringerová *et al.* 2005).

13 Potential value-added end-products obtained from kraft pulp, which is currently used  
14 mainly as paper grade pulp, are polymeric isolated hemicelluloses (Krogerus and  
15 Fuhmann 2009, Talja *et al.* 2009) and oligosaccharides (Rydlund and Dahlman 1997. Up  
16 to 60% of the xylan present in bleached hardwood market pulp can be isolated by  
17 alkaline extraction followed by precipitation and ultrafiltration (Talja *et al.* 2009). Xylan  
18 can also be isolated from the kraft cooking liquor (Dahlman *et al.* 2007) or from pulp  
19 prior to bleaching. Cold caustic extraction was used by Gomes *et al.* (2011) to remove  
20 over 60% of xylan from unbleached eucalyptus kraft pulp. In comparison to sulphite  
21 pulps higher molecular weight xylans can be obtained from kraft pulp (Janzon *et al.*  
22 2008).

1           Xylooligosaccharides (XO) are already available especially on the Asian markets  
2 for use as food ingredients to stimulate the growth of beneficial bacteria in the intestinal  
3 tract (Vázquez *et al.* 2000). XOs are produced from xylan rich feed-stocks from  
4 agriculture such as corn cobs and hulls but also wood based raw materials have been  
5 considered (Moure *et al.* 2006). XOs can be manufactured by restricted acid hydrolysis,  
6 enzymatic hydrolysis, or hydrothermal treatment either directly or after fractionation of  
7 the feedstock (Vázquez *et al.* 2000). The advantage of enzymatic hydrolysis over acid  
8 hydrolysis is specificity and thus, although the process is slower, XOs with desired  
9 degree of polymerization (DP) are obtained without the formation of monosaccharides or  
10 furfural (Akipnar *et al.* 2010).

11           With hydrolytic enzymes i.e. hemicellulases and cellulases different carbohydrate  
12 components from lignocellulosic raw materials and pulps can be selectively degraded.  
13 The selectivity of enzymatic treatment makes it an interesting process step when  
14 designing new process concepts. Several enzyme applications have been described for  
15 pulp and paper industry processes (Viikari *et al.* 2009). For example, up-grading  
16 hardwood kraft pulps into dissolving pulps has been successful by combining alkaline  
17 extraction and enzymatic treatment steps (Ibarra *et al.* 2009; Köpcke *et al.* 2010). Paper  
18 grade birch kraft can be up-graded into dissolving grade pulps by two subsequent alkali  
19 extraction steps to decrease pulp xylan content followed by endoglucanase treatment to  
20 increase pulp reactivity (Köpcke *et al.* 2010). Similar results have been obtained with  
21 eucalyptus kraft pulp by xylanase-alkaline extraction-endoglucanase sequence (Ibarra *et*  
22 *al.* 2009). The utilization of the alkaline extract obtained from such process as a source of  
23 xylan to produce xylose has also been considered (Hyatt *et al.* 1998).

1           Enzymatic treatments have separately been shown to have potential for the  
2 production of oligosaccharides from agricultural residues and for upgrading paper grade  
3 pulp into dissolving grade pulp. Based on the results by Talja *et al.* (2009), Ibarra *et al.*  
4 (2009) and Köpcke *et al.* (2010), the combination of alkaline extraction and enzymatic  
5 treatments causes formation of filtrates that contain 10 to 20% of the initial pulp  
6 carbohydrates. In order to make the process economically and environmentally feasible,  
7 these filtrates need to be exploited. However, so far the effect of the enzyme treatments  
8 on the DP of the carbohydrates present in these filtrates is not known. The possibility to  
9 obtain polymeric xylan and XOs as well as hemicellulose poor pulp from hardwood kraft  
10 pulp was evaluated in this study. The emphasis was on characterization of the effect of  
11 enzyme treatment on alkaline extraction and the effect on the molecular weight  
12 distribution of the extracted xylan as well as clarifying the possibility to isolate  
13 oligosaccharides from the pulp filtrates. In addition, the effect of enzyme treatments on  
14 the molecular weight distribution of kraft pulp polymers was clarified. Combination of  
15 xylanase or endoglucanase treatment and alkaline extraction of xylan were carried out  
16 and mass balance of the overall process was calculated.

17

## 18       **2. Material and methods**

19

### 20       **2.1. Materials**

21

22       Enzyme treatments were carried out with xylanase (pI 9) purified from *Trichoderma*  
23 *reesei* culture filtrates as described by Tenkanen *et al.* (1992). Endoglucanase treatments

1 were carried out with endoglucanase II (Cel5A) purified from *Trichoderma reesei* culture  
2 filtrate (Pere *et al.* 1995) (*Tr.* EG II) and with commercial endoglucanase product  
3 FibreCareR (Novozymes AS), which contains EG V from *Humicola insolens* (*Hi.* EG V).  
4 Xylanase and endoglucanase activities were determined as described by Pere *et al.* (1995)  
5 and the activities were expressed as katal. One nanokatal (nkat) of enzyme catalyses the  
6 release of 1 nmol of reducing sugars from the substrate polymer (birch xylan for xylanase  
7 and hydroxyethylcellulose (HEC) for endoglucanase) in one second. Protein content of  
8 the enzyme preparates was determined with BIORAD protein assay. Xylanase and  
9 endoglucanase (HEC) activities and the protein content of the used enzyme preparates are  
10 presented in Table 1.

11 Bleached commercial hardwood kraft pulp (Södra Gold Birch Z) was utilized as raw  
12 material. Before enzyme treatments or alkaline extractions dry pulp was soaked overnight  
13 in water and disintegrated with Lorentz & Wettre pulp disintegrator in 60 g batches at  
14 0.2% consistency for 30 000 revolutions.

15

## 16 **2.2. Enzyme treatments**

17

18 Enzyme treatments of pulp were carried out prior to one or in between two  
19 subsequent alkaline extraction steps. Pulp pH was adjusted to 5 with 0.5 M H<sub>2</sub>SO<sub>4</sub>.  
20 Xylanase treatment with xylanase dosage of 20 and 1000 nkat/g of dry pulp was carried  
21 out at 4% consistency, 45°C and pH 5 for 2 hours. Endoglucanase treatments with the  
22 enzyme dosage of 0.5 mg protein/g of dry pulp were carried out at 4% consistency, 45°C  
23 and pH 5 for 2 or 24 hours. Reference pulp was treated in the same way as described

1 above for 2 hours but without enzyme addition. During the enzyme treatments the pulp  
2 was mixed at 110-120 rpm. After the treatment, the pulp was heated to 90°C for 15 min  
3 to inactivate the enzymes, and thereafter it was filtered with wire cloth and washed twice  
4 with 10 ml distilled water per g of pulp.

5

### 6 **2.3. Alkaline extraction**

7

8 Alkaline extraction of pulp was carried out as described by Talja *et al.* 2009 with  
9 1 M NaOH at 5.5% consistency and at room temperature for 2 hours with mixing at 70  
10 rpm. After the alkaline extraction the pulp was filtered through wire cloth and washed  
11 thoroughly to remove the alkali. The alkaline extract contained the alkaline extract  
12 combined with the filtrate from the first washing step with 10 ml distilled water per g of  
13 pulp.

14

### 15 **2.4. Determination of carbohydrate composition**

16

17 To determine the carbohydrate composition of the pulps, pulp filtrates and  
18 extracted xylans the samples were hydrolyzed with sulphuric acid and analysed according  
19 to Willför *et al.* 2009. Pulp samples were ground with Fritch pulverisette to pass 0.5 mm  
20 screen prior to acid hydrolysis. The resulting monosaccharides were determined by  
21 HPAEC with pulse amperometric detection (Dionex ICS 3000A) equipped with  
22 CarboPac PA1 column. The polysaccharide content in the samples was calculated from  
23 the corresponding monosaccharides using an anhydro correction of 0.88 for pentoses and

1 0.9 for hexoses. Linear oligosaccharides present in the pulp filtrates were analysed by  
2 HPAEC with pulse amperometric detection (Dionex ICS 3000A) equipped with  
3 CarboPac PA1 column without the acid hydrolysis (Tenkanen et al. 1997). Linear XOs,  
4 xylobiose, xylotriose, xyloetraose, xylopentaose and xylohexaose (Megazyme) as well as  
5 cellobiose (Serva), cellotriose (Seikagaku), cellotetraose (Merck), cellopentaose  
6 (Seikagaku) and cellohexaose (Seikagaku) were used as standards. The mass balance of  
7 enzyme treatment and alkaline extraction was calculated as percentage of the original,  
8 un-extracted pulp.

9

## 10 **2.5. Determination of molecular weight of extracted xylan and pulps**

11

12 Molar mass measurements of xylan were performed by size-exclusion  
13 chromatography (SEC) in 0.1M NaOH using PSS's MCX 1000 and 100000 columns  
14 with a precolumn (0.5 ml/min, T=30°C). For pulp molar mass measurements, the  
15 extracted pulp samples were dissolved in DMAc/8% LiCl according to the solvent  
16 exchange method with ethylisocyanate derivatisation (Berthold *et al.* 2001). The SEC  
17 measurements were performed using 2 x PL gel MiniMixed A columns with a precolumn  
18 in DMAc/0.8% LiCl eluent (0.36 ml/min, T=80 °C). In both cases, the refractive index  
19 (RI) detector was used, and the molar mass distributions of xylan and pulp  
20 polysaccharides were calculated in relation to pullulan standards using Waters Empower  
21 2 software.



1

## 2       **3. Results and discussion**

3

### 4       **3.1. Enzyme hydrolysis efficiency in the original and alkali extracted pulp**

5

6       Efficiency and specificity of the enzymatic hydrolysis of the bleached hardwood  
7 (HW) kraft pulp before and after alkaline extraction was quantified based on  
8 carbohydrates released from the pulp in the enzyme treatment. The analysis of the  
9 carbohydrates was carried out from the pulp filtrates after acid hydrolysis to  
10 monosaccharides. It was noted from the carbohydrates released to pulp filtrate that the  
11 treatments with xylanase and *Humicola insolens* EG V were quite specific towards xylan  
12 and cellulose, respectively (Fig. 1). *Trichoderma reesei* endoglucanase II released some  
13 xylose to the filtrate, which is probably caused by the minor xylanase impurity in the  
14 preparate (Table 1). Xylanase treatment (1000 nkat/g) hydrolyzed 2.9% of the un-  
15 extracted, original hardwood kraft pulp and only 1.1% of the pulp after the alkaline  
16 extraction (Fig. 1). The alkali-extracted pulp was hydrolyzed more efficiently than the  
17 un-extracted pulp by the both endoglucanases (Fig. 1).

18       It is well known that the enzyme reactions happen on the fibre surfaces and fines as  
19 the molecular size of the enzymes (5 to 10 nm diameter) hinders their penetration deep  
20 into the fibres (Suurnäkki *et al.* 1996). Xylan of the bleached HW kraft fibres is enriched  
21 at the fibre surface (Dahlman *et al.* 2003) and in fines (Lyytikäinen *et al.* 2011). Thus, it  
22 can be speculated that xylan at the surface of un-extracted HW kraft pulp fibres is easily  
23 accessible to xylanase but hinders the hydrolysis by EGs. In addition, alkaline extraction

1 increases the fibre porosity (Lyytikäinen *et al.* 2011), which enhances the penetration of  
2 the enzymes into the fibres and may have affected the hydrolysis efficiency of alkali-  
3 extracted pulp by xylanase.

4         The extensive removal of xylan and the subsequently increased fibre porosity by  
5 alkaline extraction apparently improved the accessibility of fibre cellulose to  
6 endoglucanases. Increased efficiency of endoglucanase hydrolysis after alkaline  
7 extraction has also been observed by Köpcke *et al.* (2010). In their study the enhanced  
8 hydrolysis was observed as increased reduction of pulp viscosity by commercial  
9 endoglucanase (Novozym 476) treatment of pulp after three alkaline extractions in  
10 comparison to pulp after two alkaline extractions. Lower molar mass of pulp  
11 polysaccharides, *i.e.* cellulose, was also detected in our experiments, when endoglucanase  
12 treatment was performed between two alkaline extractions (Fig. 3, Table 2).

13

### 14         **3.2. Release of oligosaccharides to pulp filtrates**

15

16         Oligosaccharides formed during the enzyme treatments of original and alkali-  
17 extracted bleached HW kraft pulp were analysed by HPAEC-PAD from the pulp filtrates.  
18 As expected, xylooligosaccharides (XO) were formed by xylanase treatment and cello-  
19 oligosaccharides by EG treatments (Fig. 2). By comparing the amount of  
20 oligosaccharides (Fig. 2) and the total amount of released carbohydrates (analysis after  
21 acid hydrolysis, Fig. 1) it can be concluded that the carbohydrates were released to the  
22 filtrate mainly as oligosaccharides. XOs with DP two to six were released by the xylanase  
23 treatment, the main two hydrolysis products being xylobiose and xylotriose, which is in

1 agreement with previous results (Tenkanen *et al.* 1992). The yield of XOs with the higher  
2 xylanase dosage (1000 nkat/g) was 12% or 3.8% of the original pulp xylan when the  
3 enzyme treatment was carried out to un-extracted or alkali-extracted pulp, respectively.  
4 The obtained DP profile of the pulp hydrolysate after xylanase treatment shows potential,  
5 as DP two to four XOs are preferred for food applications (Vázquez *et al.* 2000).

6 The cello-oligosaccharide profile released by the two endoglucanases differed  
7 from each other. *Trichoderma reesei* EGII released cellobiose and cellotriose as well as  
8 glucose, whereas *Humicola insolens* released longer oligosaccharides, mainly cellobiose,  
9 cellotriose and cellotetraose from the pulp.

10

### 11 **3.3. Analysis of extracted xylans**

12

13 Carbohydrate composition of alkaline extracts from enzyme aided xylan extraction  
14 was determined after pH adjustment and acid hydrolysis with HPAEC-PAD (Table 2).  
15 When endoglucanase treatment preceded the alkaline extraction also glucose was present  
16 in the alkaline extracts, whereas other xylan samples were of high purity. These results,  
17 carried out with bleached HW pulp comprising mainly of birch, are in accordance with  
18 the previously reported results showing that high purity (98 – 99% carbohydrates) xylan  
19 can be obtained from bleached birch pulp (Talja *et al.* 2009; Janzon *et al.* 2008).

20 The xylan content in alkaline extracts derived from the first extraction of the pulp  
21 was tenfold higher than that from the second extraction step (Table 2). Xylan yield of the  
22 reference pulp in the first extraction was 61% of the original pulp. Xylanase treatment  
23 decreased the extracted xylan yield by up to 10%, which corresponds to the yield of

1 xylooligosaccharides (XOs) in the pulp filtrate (Table 2, Fig. 2). The extraction of xylan  
2 was not affected by the endoglucanase treatment, suggesting that the enzyme treatment  
3 has not improved the penetration of the alkali or altered the binding of xylan to cellulose.

4 After the second alkaline extraction of pulp, 7% of xylan present in the original un-  
5 extracted pulp was observed in the extract. The effects of the enzyme pre-treatments on  
6 the xylan yield in the second extraction, varying from 7 to 9% of original pulp xylan,  
7 were only minor.

8 Molar mass distributions of the isolated, alkali-extracted, xylans were determined  
9 by SEC in 0.1M NaOH (Table 2). In the reference case the xylan extracted in the second  
10 stage had somewhat higher molar mass than that extracted in the first stage. Xylanase  
11 treatment decreased the molecular mass of the isolated xylan in both extraction stages,  
12 although the effect was more pronounced in the second stage. This suggests that pulp  
13 xylan after first alkaline extraction was more efficiently hydrolyzed by the xylanase  
14 treatment in comparison to the un-extracted pulp.

15 The observed decrease in the molecular weight of xylan isolated from xylanase  
16 treated pulp was expected. A decrease in the molecular weight of xylan extracted from  
17 xylanase treated O-delignified eucalyptus kraft pulp was observed also by Gehmayr and  
18 co-workers (2011). However, the used xylanase dosage was apparently higher than that  
19 used in our work as even 46% of pulp xylan was hydrolyzed and xylan Mw was 50%  
20 lower than that of the reference.

21 Endoglucanase treatment had smaller effect on xylan molar mass. It should be noted  
22 that the *T. reesei* EG II preparate contained also minor xylanase activity (Table 1),  
23 probably causing the observed decrease in xylan Mw observed especially in the second

1 extraction after 24h treatment. *H. insolens* EG V was specific towards cellulose and did  
2 not decrease the molar mass of extracted xylan even after long incubation time. An  
3 additional low molecular weight (Mw ~3 500 Da) fraction was observed in the extracts  
4 derived from EG V treated pulps (results not shown). This fraction probably is derived  
5 from cellulose degradation products from enzymatic hydrolysis.

6         The molar mass distribution obtained in our work is comparable to that obtained  
7 by Talja *et al.* 2009. However, considerably lower molar mass xylan, Mw 11 000 D was  
8 obtained by Janzon *et al.* 2008 from birch kraft pulp by using KOH, NaOH and nitren  
9 extractions. This supports the suggestion by Janzon *et al.* 2008, that DP of xylans  
10 extracted from pulps depend on the pulping conditions and thus differences in Mw of  
11 xylans isolated from different pulps are expected. It should also be noted, that the molar  
12 mass detection method used here is relative, and very much dependent on used measuring  
13 conditions, e.g. used eluent.

14

### 15         **3.4. Properties of alkali-extracted pulps**

16

17         Composition of kraft pulp samples from the sequential alkaline extraction and  
18 enzyme treatments are presented in Table 3. Pulp xylan content was 10 to 12% after the  
19 first and 6 to 7% after the second alkali-extraction stage. The decrease in xylan content in  
20 xylanase treated samples is comparable to the amount of xylan hydrolyzed during the  
21 enzyme treatment prior to the first alkaline extraction (Table 2, Fig. 1). After the second  
22 alkaline extraction only minor differences were observed in the composition of the  
23 reference and enzyme treated pulp samples.

1           Analysis of pulp molar mass distribution after different treatments indicated that  
2 the alkaline extraction of xylan prior to the endoglucanase treatment enhances the  
3 hydrolytic effect of endoglucanases on pulp cellulose (Fig. 3, Table 3). Based on the pulp  
4 molar mass distribution the cellulose hydrolysis was more efficient with *H. insolens* EG  
5 V compared to *T. reesei* EG II and the difference is even more emphasized when EG  
6 treatment is performed after alkaline extraction (Fig. 3). In addition the pulp molar mass  
7 distribution was narrower after the treatment with *H. insolens* EG V than *T. reesei* EG II.  
8 The higher reduction of cellulose molar mass by endoglucanases between alkaline  
9 extractions is well in accordance with the results of Köpcke *et al.* (2010). The xylanase  
10 treatments had no effect on molar mass distribution of pulp polysaccharides analysed  
11 after alkaline extractions.

12  
13

### 14           **3.5. Mass balance of enzyme-aided xylan extraction**

15

16           The mass balance of the enzyme aided alkaline extraction of birch kraft pulp with  
17 one or two subsequent alkaline extraction steps are presented in Table 4. The total yield  
18 comprises of the extracted pulp, oligosaccharides and other carbohydrates analyzed from  
19 the pulp filtrate, and xylan and other carbohydrates analyzed from the alkaline extract.  
20 When the enzyme treatment was carried out between two subsequent alkaline extraction  
21 steps the mass balance was calculated as percentage from the original un-extracted pulp,  
22 making the theoretical overall yield 85%. The total yield of the first alkaline extraction  
23 step varied from 90 to 101%. The low 90% overall yields were observed for pulps with

1 the most extreme enzyme treatment, either high xylanase dosage or long incubation time  
2 with endoglucanase. The possible explanation for the low overall yield observed in these  
3 cases is that some of the pulp material has been lost during pulp washing steps. In the  
4 treatment of alkali-extracted pulp the total yields are close to 100% of the theoretical  
5 maximum.

6 The xylan yield of the reference sample in the first alkaline extraction stage was 15%  
7 of the original pulp and 61% the original pulp xylan. The xylan yield was decreased by  
8 the extensive xylanase treatment to 12% of the original pulp, with simultaneous 2.8%  
9 yield of xylooligosaccharides (XO) in the pulp filtrate (Table 4). In the two stage alkali-  
10 extraction, the extracted xylan yield after the second extraction step was about 2% of the  
11 original pulp 7-9% of the xylan present in the original un-extracted pulp depending on the  
12 enzyme treatment used (Tables 3 and 4).

13

#### 14 **4. Conclusions**

15

16 Hemicellulose poor pulps as well as oligosaccharides and polymeric xylan were  
17 obtained by combining specific enzymatic treatments and alkaline extraction. Sequential  
18 alkaline extraction of hardwood kraft pulp yielded two high purity xylan fractions, one  
19 with high yield (60% of original pulp xylan) and one with high molecular weight (up to  
20 40 000 Da) but considerably lower yield (~7% of original pulp xylan). Reduction of  
21 cellulose DP by both *T. reesei* EG II and *H. insolens* EG V was significantly enhanced  
22 after xylan removal.

1 Xylanase treatment prior to alkaline extraction decreased the yield and molecular  
2 weight of extracted xylan but provided the possibility to isolate xylooligosaccharides  
3 (~10% of original xylan) as an additional value added component. To obtain xylan with  
4 un-altered molecular weight as well as xylooligosaccharides (3.8% of original xylan),  
5 xylanase treatment should be carried out after alkaline extraction of xylan. Thus,  
6 depending on the desired xylan fraction, the xylanase treatment should be omitted  
7 (polymeric xylan with high DP obtained), carried out before the alkaline extraction  
8 (xylooligosaccharides and polymeric xylan with decreased DP obtained) or after the  
9 alkaline extraction (polymeric xylan with high DP obtained and xylooligosaccharides  
10 obtained).

11 The effects of EG treatments on the pentose containing filtrates were minor and xylan  
12 extracts comparable to that from reference were obtained after EG treatments. Thus EG  
13 treatment does not negatively affect the extraction of polymeric xylan. The main benefits  
14 obtained by EG treatment lies in its positive effect on the pulp reactivity (Köpcke et al.  
15 2010) and DP. In our experiments *H.insolens* EG V decreased the pulp polydispersity,  
16 which is a desired property for a dissolving grade pulp.

17 It is thus possible to utilize similar process sequence than previously used for  
18 upgrading hardwood kraft pulp into dissolving pulp for obtaining both polymeric xylan  
19 and XOs together with hemicellulose lean hardwood kraft pulp.

20

21 **Acknowledgments**

22



1 Teija Jokila, Nina Vihersola and Eila Turunen are acknowledged for skilful technical  
2 assistance. Atte Mikkelsen is thanked for assistance in mono- and oligosaccharide  
3 analysis. Hanne Host-Pedersen (Novozymes A/S) is thanked for providing the  
4 commercial enzyme product. The research leading to these results has received funding  
5 from the European Community's Seventh Framework Programme FP7/2007-2013 under  
6 grant agreement n° CP-IP 228589-2 AFORE.

7

## 8 **References**

9

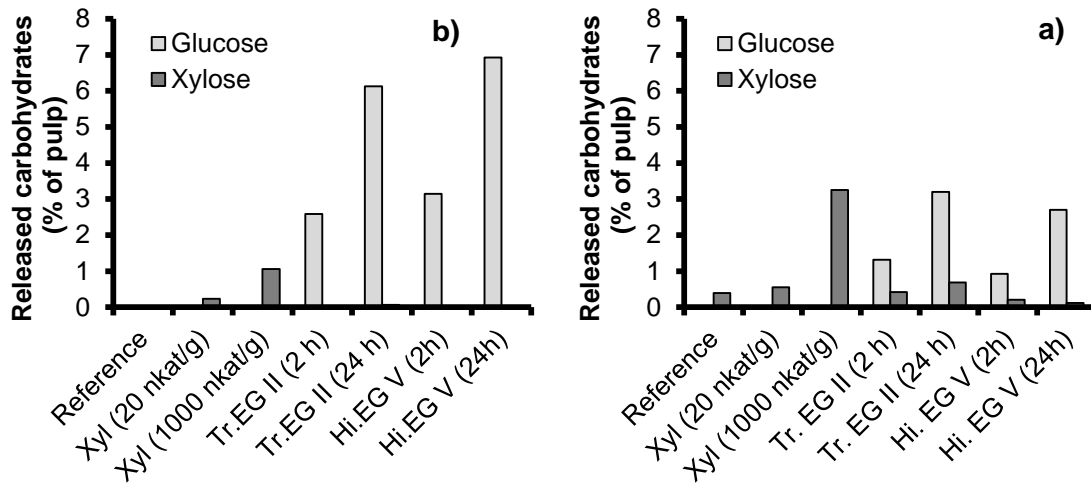
- 10 Akipnar, O., Erdogan, K., Bakir, U., & Yilmaz L. (2010). Comparison of acid and  
11 enzymatic hydrolysis of tobacco stalk xylan for preparation of xylooligosaccharides.  
12 *LWT – Food Sci Technol.*, 43, 119-125.
- 13 Berthold, F., Gustafsson, K., Sjöholm, E., & Lindstrom, M. (2011). An improved method  
14 for the determination of softwood kraft pulp molecular mass distributions, 11<sup>th</sup>  
15 International Symposium on Wood and Pulping Chemistry, Nice, France, June 11-14,  
16 Vol 1, 363-366.
- 17 Dahlman, O., Jacobs, A., & Sjöberg, J. (2003). Molecular properties of hemicelluloses  
18 located in the surface and inner layers of hardwood and softwood pulps. *Cellulose*,  
19 10, 325-334.
- 20 Dahlman, O., Tomani, P., Axegård, P., Lundqvist, P., & Lindgren, K. (2007). Method for  
21 separating polymeric pentose from a liquid/slurry. WO/2007/120091
- 22 Ebringerová, A., Hromádková, Z., & Heinze, T. (2005). Hemicellulose. *Adv Polym Sci*,  
23 186, 1–67.

- 1 Gehmayr, V., Schild, G., & Sixta, H. (2011). A precise study on the feasibility of enzyme  
2 treatments of a kraft pulp for viscose application. *Cellulose* 18, 479-491.
- 3 Gomes, V.J., Colodette, J.L., Barbosa, L.C.A. & Oliveira, R.C (2011) Improved methods  
4 for dissolution and tissue grade paper pulps production from eucalypt. Proceedings of  
5 Italic 6 - Science and technology of biomasses: Advances and challenges, September  
6 5-8, 2011, Viterbo, Italy, pp. 99-103.
- 7 Hyatt, J., Fengl, R.W., Edgar, K.J., & Alvarz-Wright, M.T. (1998). Process for the co-  
8 production of dissolving pulp and xylan. International patent application  
9 WO98/16682.
- 10 Ibarra, D., Köpcke, V., & Ek, M. (2009). Exploring enzymatic treatments for the  
11 production of dissolving grade pulp from different wood and non-wood paper grade  
12 pulps. *Holzforschung*, 63, 721-730.
- 13 Janzon, R., Saake, B., & Puls J. (2008). Upgrading of paper-grade pulps to dissolving  
14 pulps by nitren extraction: properties of nitren extracted xylans in comparison to  
15 NaOH and KOH extracted xylans. *Cellulose*, 15, 161-175.
- 16 Krogerus, B. & Fuhrmann, A. (2009) Isolation of xylan and use as wet-end and binder  
17 chemical. Proceedings of 15<sup>th</sup> International Symposium on Wood, Fibre and Pulping  
18 Chemistry. June 15-18, 2009, Oslo, Norway.
- 19 Köpcke, V., Ibarra, D., Larsson, P.T, & Ek, M. (2010). Optimization of treatment  
20 sequences for the production of dissolving pulp from birch kraft pulp. *NPPRJ* 25, 31-  
21 38.

- 1 Lyytikäinen, K., Saukkonen, E., Kajanto, I., & Käyhkö, J. (2011). The effect of  
2 hemicellulose extraction on fiber charge properties and retention behavior of kraft  
3 pulp fibers. *BioRes.*, 6, 219-231.
- 4 Moure, A., Gullón, P., Domínguez, H. & Parajo, J.C. (2006). Advances in the  
5 manufacture, purification and applications of xylo-oligosaccharides as food additives  
6 and nutraceuticals. *Process Biochemistry*, 41, 1913–1923.
- 7 Pere, J., Siika-aho, M., Buchert, J., & Viikari, L. (1995). Effects of purified *Trichoderma*  
8 *reesei* cellulases on the fiber properties of kraft pulp. *Tappi J*, 78, 71-78.
- 9 Rydlynd A. & Dahlman, O. (1997). Oligosaccharides obtained by enzymatic hydrolysis  
10 of birch kraft pulp xylan: Analysis by capillary zone electrophoresis and mass  
11 spectrometry. *Carbohydr. Res.*, 300, 95-102.
- 12 Talja, R.A., Fuhrman, A., Krogerus, B., & Vähä-Nissi, M. (2009). Hemicelluloses from  
13 biomass to applications. Proceedings of the 7th biennial Johan Gullichsen colloquium,  
14 2009, 77-84.
- 15 Tenkanen, M., Makkonen, M., Perttula, M., Viikari, L., & Teleman, A. (1997). Action of  
16 *Trichoderma reesei* mannanase on galactoglucomannan on pine kraft pulp. *J.*  
17 *Biotechnol.*, 57, 191-204.
- 18 Tenkanen, M., Puls, J., & Poutanen, K. (1992). Two major xylanases of *Trichoderma*  
19 *reesei*. *Enzyme Microb. Technol.*, 14, 566-574.
- 20 Suurnäkki, A., Heijnesson, A., Buchert, J., Tenkanen, M., Viikari, L. & Westermark, U.  
21 (1996). Location of xylanase and mannanase action in kraft fibers. *JPPS*, 22, J78-J83.
- 22 Vázquez, M.J., Alonso, J.L., Domínguez, H. & Parajo, J.C. (2000).  
23 Xylooligosaccharides: manufacture and applications. *Trends Food Sci Technol*, 11,  
24 387-393.

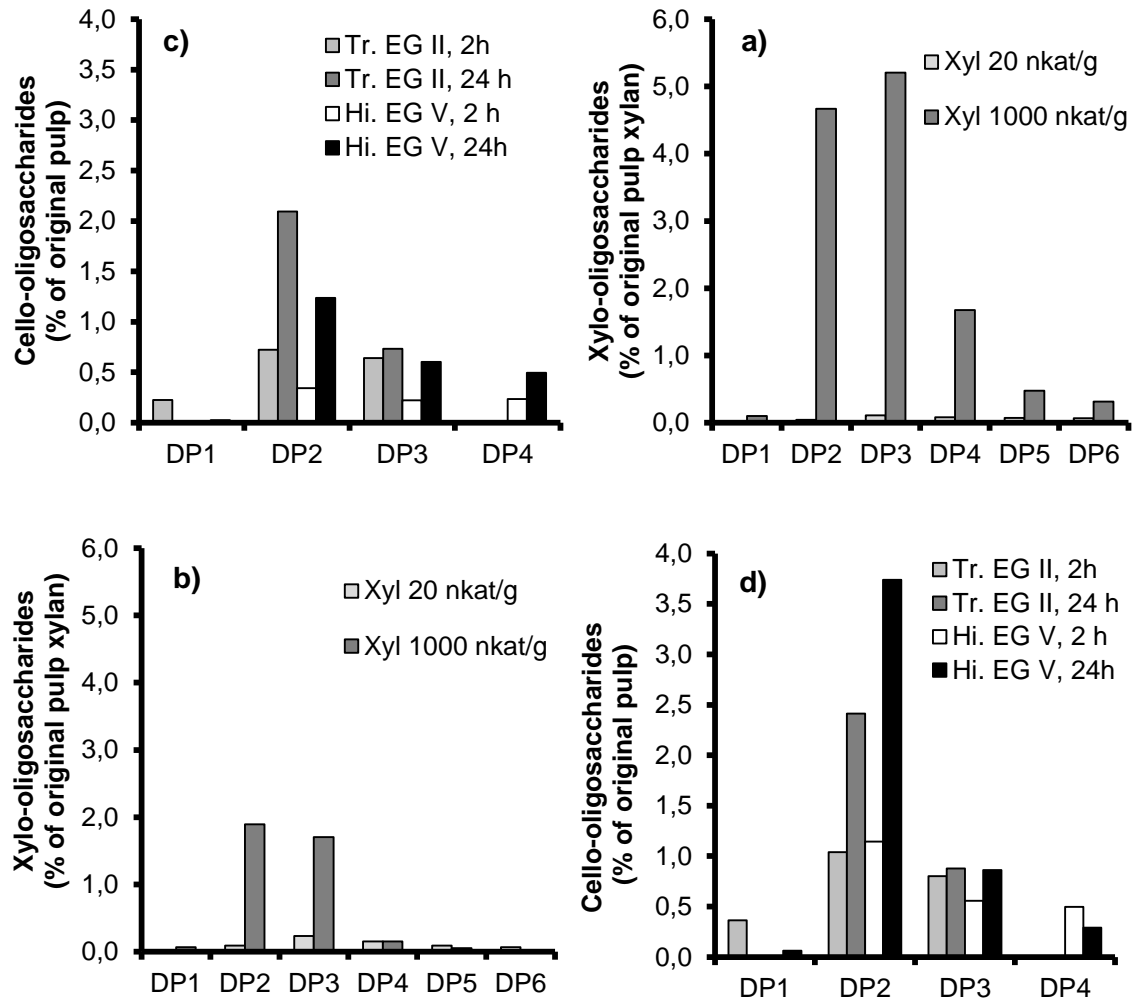
- 1 Viikari, L., Suurnäkki, A., Grönqvist, S., Raaska, L., & Ragauskas, A. (2009). Forest  
2 Products: Biotechnology in Pulp and Paper Processing, In: Encyclopedia of  
3 Microbiology. 3<sup>rd</sup> ed. Schaechter, M. (ed.). Academic Press, pp. 80 – 94.
- 4 Willför, S., Pranovich, A., Tamminen, T., Puls, J., Laine, C., Suurnäkki, A., Saake, B.,  
5 Uotila, K., Simolin, H., Hemming, J. & Holmbom, B. (2009). Carbohydrate analysis  
6 of plant materials with uronic acid-containing polysaccharides–A comparison  
7 between different hydrolysis and subsequent chromatographic analytical techniques.  
8 *Ind crops prod.*, 29, 571–580.
- 9

1 **Figures:**



2

3 **Figure 1.** Carbohydrates released from the un-extracted (a) and once alkali-extracted (b)  
 4 bleached HW kraft pulp in the enzyme treatment. Analysis was carried out after acid  
 5 hydrolysis by HPAEC and calculated as % of the dry pulp in the treatment. Reference: no  
 6 enzyme addition; Xyl: *Trichoderma reesei* pI 9 xylanase, 2 h; Tr. EGII: *Trichoderma*  
 7 *reesei* EG II (Cel 5A) 0.5 mg/g pulp; Hi. EG V: *Hemicola insolens* EG V (FibreCareR)  
 8 0.5 mg/g pulp.

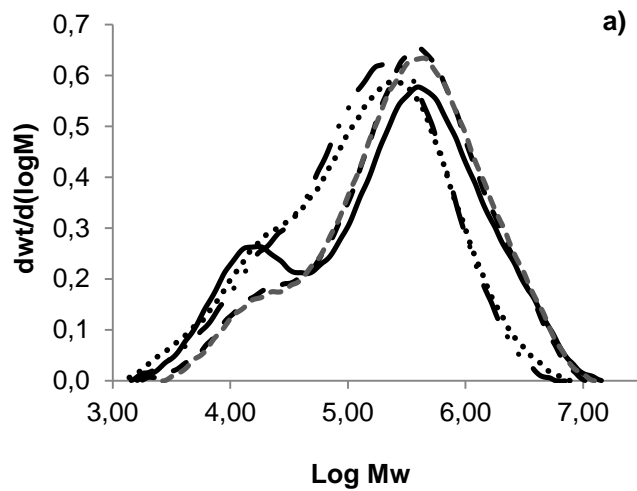


1

2

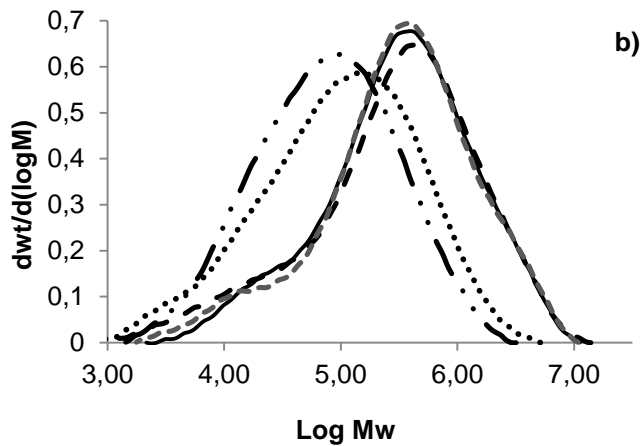
3 **Figure 2.** Mono- and oligosaccharides formed during the enzyme treatment of un-  
 4 extracted and alkali-extracted HW Kraft pulp. Xylooligosaccharides (XO) released  
 5 during xylanase treatment of un-extracted (a) and extracted (b) pulp and cello-  
 6 oligosaccharides formed during EG treatments of un-extracted (c) and extracted (d) pulp.  
 7 Reference: no enzyme addition; Xyl: *Trichoderma reesei* pI 9 xylanase, 2 h; Tr. EGII:  
 8 *Trichoderma reesei* EG II (Cel 5A) 0.5 mg/g pulp; Hi. EGV: *Humicola insolens* EG V  
 9 (FibreCareR) 0.5 mg/g pulp.

10



— Raw material to 1st extr  
 - - Reference  
 - · - Xyl 1000 nkat  
 ····· Tr. EG II 24h  
 - · - Hi. EG V 24h

1



— Raw material to 2nd Extr  
 - - Reference  
 - · - Xyl 1000 nkat  
 ····· Tr. EG II 24h  
 - · - Hi. EG V 24 h

2

3 **Figure 3.** Effect of enzyme treatments on molar mass distributions of the pulp  
 4 polysaccharides after the first (a) and the second (d) alkaline extraction. Reference: no  
 5 enzyme addition; Xyl: *Trichoderma reesei* pI 9 xylanase, 2 h, 1000 nkat; Tr. EG II:  
 6 *Trichoderma reesei* EG II (Cel 5A) 0.5 mg/g pulp, 24 h; Hi. EG V: *Humicola insolens*  
 7 EG V (FibreCareR) 0.5 mg/g pulp, 24 h.

8

1 **Table 1.** Enzyme activities and protein content of the used enzyme preparates

	Xylanase (nkat/ml)	HEC <sup>a</sup> (nkat/ml)	Protein (mg/ml)
<i>Trichoderma reesei</i> , Xylanase, pI 9	31 000	ND <sup>b</sup>	3.2
<i>Trichoderma reesei</i> , EG II (Cel5a)	11.5	6 886	7.5
<i>Humicola insolens</i> , EG V	ND	1 520	12.0

2 <sup>a</sup>HEC: Hydroxyethylcellulose

3 <sup>b</sup>ND: Not detected

4

5 |



1 **Table 2.** Composition, yield and average molar mass (Mn, Mw) of alkaline extracts from  
 2 enzyme aided xylan extraction of HW Kraft Pulp.

Treatment sequence <sup>a</sup>	Content in alkaline extract (%)		Average molar masses			Xylan yield (%) <sup>b</sup>
	Xylan	Glucose	Mn	Mw	PD	
First extraction stage						
Reference – Alk	5.3	ND <sup>c</sup>	27 400	36 300	1.3	61
Xylanase 20 nkat – Alk	4.8	ND	25 200	34 900	1.4	55
Xylanase 1000 nkat – Alk	4.4	ND	16 600	24 700	1.5	50
<i>Tr.</i> EG II, 2h – Alk	5.0	0.05	25 800	35 300	1.4	58
<i>Tr.</i> EG II, 24 h – Alk	4.9	0.05	23 900	32 900	1.4	56
<i>Hi.</i> EG V 2 h – Alk	5.7	0.07	26 700	36 700	1.4	64
<i>Hi.</i> EG V 24 h – Alk	5.1	0.07	26 800	36 600	1.4	58
Second extraction stage						
Alk - Reference – Alk	0.6	ND	31 000	41 300	1.3	7.2
Alk - Xylanase 20 nkat – Alk	0.7	ND	19 700	29 300	1.5	8.3
Alk - Xylanase 1000 nkat – Alk	0.7	ND	12 700	20 000	1.6	8.0
Alk - <i>Tr.</i> EG II, 2h – Alk	0.8	0.1	31 800	42 000	1.3	9.0
Alk - <i>Tr.</i> EG II, 24 h – Alk	0.8	0.1	26 500	35 700	1.4	9.6
Alk - <i>Hi.</i> EG V 2 h – Alk	0.6	0.2	31 700	41 200	1.3	7.3
Alk - <i>Hi.</i> EG V 24 h – Alk	0.7	0.2	30 900	40 100	1.3	8.2

3 <sup>a</sup>Reference: no enzyme addition, Xylanase: *Trichoderma reesei* pI 9 xylanase, 2 h; *Tr.* EGII: *Trichoderma*  
 4 *reesei* EG II (Cel 5A) 0.5 mg/g pulp; *Hi.* EGV: *Humicola insolens* EG V (FibreCareR) 0.5 mg/g pulp. Alk:  
 5 Alkaline extraction: 1 M NaOH at 5.5% consistency, room temperature, 2 hours

6 <sup>b</sup>Xylan yield as % of xylan present in original, un-extracted pulp

7 <sup>c</sup>ND: Not detected, detection limit 4 mg/l

8

1 **Table 3.** Composition, average molar mass and yield of alkali-extracted pulps with or  
 2 without enzyme treatment.

Treatment sequence <sup>a</sup>	Xylan (%)	Cellulose (%)	Other carboh. (%)	Pulp yield (%) <sup>b</sup>	Average molar masses		
					Mn (g/mol)	Mw (g/mol)	PD
Raw material to 1 <sup>st</sup> extraction stage experiments	23.6	75.9	0.5	100	42 300	728 900	17.2
Reference – Alk	12.0	87.5	0.5	83	67 900	721 000	10.6
Xylanase 20 nkat – Alk	11.3	88.3	0.5	82	64 800	725 600	11.2
Xylanase 1000 nkat – Alk	10.6	88.9	0.5	75	77 000	775 000	10.1
<i>Tr.</i> EG II, 2h – Alk	12.4	87.1	0.5	78	46 400	469 700	10.1
<i>Tr.</i> EG II, 24 h – Alk	12.7	86.8	0.5	80	34 800	377 200	10.8
<i>Hi.</i> EG V 2 h – Alk	10.2	89.4	0.4	84	47 100	397 100	8.4
<i>Hi.</i> EG V 24 h – Alk	10.6	89.0	0.5	73	41 800	339 700	8.1
Raw material to 2 <sup>nd</sup> extraction stage experiments	9.3	90.2	0.5	85	95 500	782 600	8.1
Alk - Reference – Alk	6.2	93.5	0.4	83	52 600	801 500	15.2
Alk - Xylanase 20 nkat – Alk	7.1	92.4	0.5	83	61 200	760 600	12.4
Alk - Xylanase 1000 nkat – Alk	6.4	93.1	0.5	82	77 000	790 600	10.2
Alk - <i>Tr.</i> EG II, 2h – Alk	7.1	92.4	0.5	81	41 900	365 400	8.7
Alk - <i>Tr.</i> EG II, 24 h – Alk	5.6	94.0	0.4	78	27 600	262 700	9.5
Alk - <i>Hi.</i> EG V 2 h – Alk	7.0	92.6	0.5	80	26 800	221 100	8.2
Alk - <i>Hi.</i> EG V 24 h – Alk	6.7	92.8	0.4	77	30 200	180 300	6.0

3 <sup>a</sup> Raw material for the 1<sup>st</sup> extraction stage is un-extracted commercial HW bleached kraft pulp, raw material  
 4 for the 2<sup>nd</sup> extraction stage is the same pulp after one alkaline extraction stage. Reference: no enzyme  
 5 addition; Xylanase: *Trichoderma reesei* pI 9 xylanase, 2 h; *Tr.* EGII: *Trichoderma reesei* EG II (Cel 5A)  
 6 0.5 mg/g pulp; *Hi.* EGV: *Humicola insolens* EG V (FibreCareR) 0.5 mg/g pulp; Alk: Alkaline extraction  
 7 with 1 M NaOH at 5.5% consistency, room temperature, 2 hours

8 <sup>b</sup>Pulp yield as % of the original, un-extracted pulp  
 9

1 **Table 4.** Mass balance of enzyme-aided first and second alkaline extraction stage of HW  
 2 Kraft Pulp as percentage of the original pulp.

Treatment sequence <sup>a</sup>	Pulp yield (%)	Oligosac. yield (%) <sup>b</sup>	Other carboh in filtrate (%)	Xylan yield (%)	Other carboh in xylan (%)	Total yield (%) <sup>c</sup>
Reference – Alk	83	ND <sup>d</sup>	0.4	15	ND	98
Xylanase 20 nkat – Alk	82	0.1	0.5	13	ND	96
Xylanase 1000 nkat – Alk	75	2.8	0.4	12	ND	90
<i>Tr.</i> EG II, 2h – Alk	78	1.4	0.4	14	0.13	94
<i>Tr.</i> EG II, 24 h – Alk	80	2.8	1.1	14	0.13	98
<i>Hi.</i> EG V 2 h – Alk	84	0.8	0.3	16	0.19	101
<i>Hi.</i> EG V 24 h – Alk	73	2.3	0.5	14	0.20	90
Alk - Reference – Alk	83	ND	ND	1.5	ND	85
Alk - Xylanase 20 nkat – Alk	83	0.1	0.1	1.7	ND	85
Alk - Xylanase 1000 nkat – Alk	82	0.8	0.1	1.7	ND	84
Alk - <i>Tr.</i> EG II, 2h – Alk	81	1.8	0.3	1.9	0.2	86
Alk - <i>Tr.</i> EG II, 24 h – Alk	78	3.3	1.9	2.0	0.3	86
Alk - <i>Hi.</i> EG V 2 h – Alk	80	2.2	0.5	1.5	0.4	85
Alk - <i>Hi.</i> EG V 24 h – Alk	77	4.9	1.0	1.7	0.5	85

3 <sup>a</sup>Reference: no enzyme addition; Xylanase: *Trichoderma reesei* pI 9 xylanase, 2 h; *Tr.* EGII: *Trichoderma*  
 4 *reesei* EG II (Cel 5A) 0.5 mg/g pulp; *Hi.* EGV: *Humicola insolens* EG V (FibreCareR) 0.5 mg/g pulp. Alk:  
 5 Alkaline extraction with 1 M NaOH at 5.5% consistency, room temperature, 2 hours

6 <sup>b</sup>Xylooligosaccharides in the case of xylanase and celooligosaccharides in the case of EG

7 <sup>c</sup>In the second extraction stage the theoretical maximal yield is 84% of original un-extracted pulp

8 <sup>d</sup>ND= Not detected, detection limit 4 mg/l

9